Supplementary data for the article:

Korać, J.; Stanković, D. M.; Stanić, M.; Bajuk-Bogdanović, D.; Žižić, M.; Pristov, J. B.; Grgurić-Šipka, S.; Popović-Bijelić, A.; Spasojević, I. Coordinate and Redox Interactions of Epinephrine with Ferric and Ferrous Iron at Physiological PH. *Scientific Reports* **2018**, 8 (1). https://doi.org/10.1038/s41598-018-21940-7

Coordinate and redox interactions of epinephrine with ferric and ferrous iron at physiological pH

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Supplementary Information

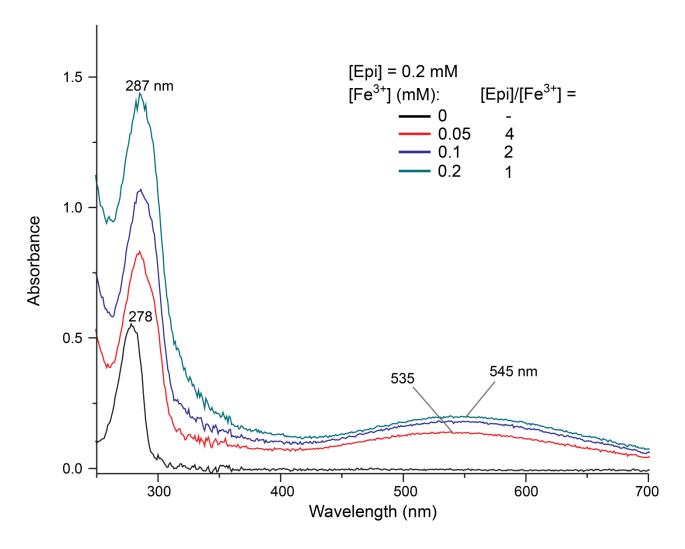


Figure S1 UV/Vis spectra of Epi and ferric iron in 10 mM phosphate buffer, pH 7.4. In contrast to Tris, the same complex appears to predominate at both low and high [Epi]/[Fe³⁺] ($\lambda_{max} = 545$ nm), most likely in relation to the high affinity of phosphates for Fe³⁺. The spectra were acquired following 15 min incubation, and remained unaltered for at least 1 h.

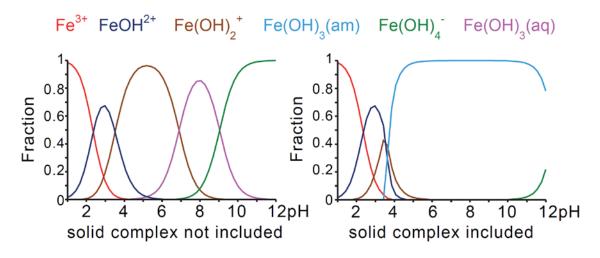


Figure S2 Speciation diagrams of Fe³⁺ in water. Diagrams were prepared in Hydra-Medusa Software, using the following parameters: $[Fe^{3+}]$ = 0.1 mM; pH range 1–12; T = 293 K.

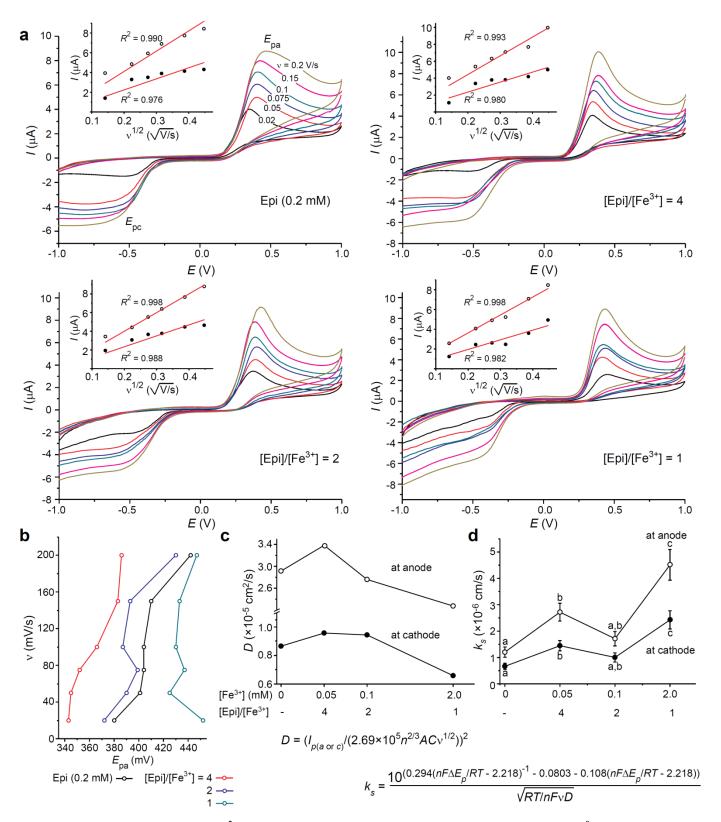


Figure S3 Scan rate analysis of the Epi/Fe³⁺ systems. (a) Cyclic voltammograms of Epi in absence or presence of Fe³⁺ at the boron doped diamond electrode obtained at different scan rates (v = 0.02-0.2 V/s). The inset figures in each panel represent the dependence between I_{pa} (open circles) and I_{pc} (closed circles; absolute values) and $v^{1/2}$. Linear fit and R^2 values are presented. (b) E_{pa} for Epi and [Epi]/[Fe³⁺] = 4, 2,

and 1 at different v. (c) D for Epi and different [Epi]/[Fe³⁺]. Randles–Sevick equation (at the bottom middle): n, number of transferred e⁻; A, area of the working electrode (0.0707 cm²); C, concentration of redox species in solution (mol/cm³). (d) Rate constants of electron transfer (k_s) for Epi and different [Epi]/[Fe³⁺]. Results are presented as means (\pm SE) of measurements made at various v. k_s not sharing a common letter are significantly different (P < 0.05). Nicholson Shain calculus (at the bottom right): R, standard gas constant; T, temperature (298 K); F, Faraday's constant; ΔE_p , the difference between E_{pa} and E_{pc} taken at various v.

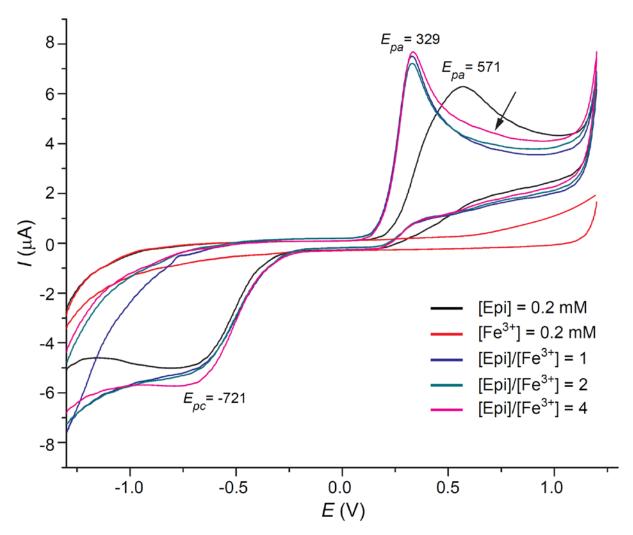


Figure S4 Cyclic voltammograms of Epi in absence or presence of Fe³⁺ in 10 mM potassium phosphate buffer, pH 7.4, at the boron doped diamond electrode. The oxidation/anodic (E_{po}) and reduction/cathodic (E_{po}) potentials are presented. Scan rate was 0.1 V/s.

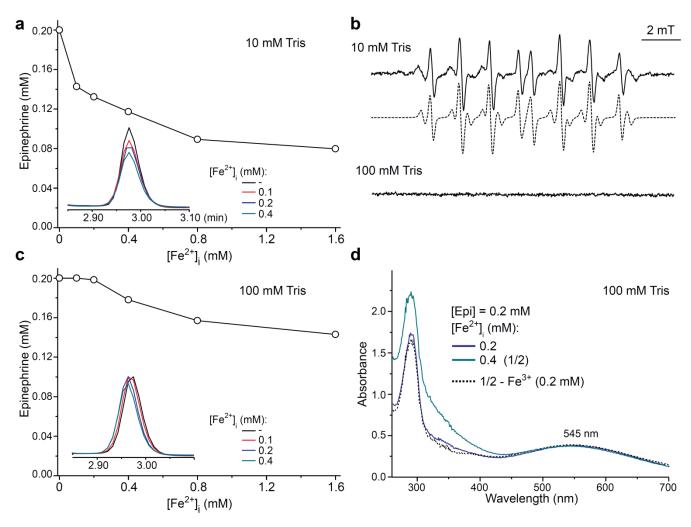


Figure S5 Antioxidative performance of 10 mM and 100 mM Tris, pH 7.4. (a) The concentration of Epi following 5 min incubation with different [Fe²⁺]_i in 10 mM Tris buffer. Inset: Epi peaks in HPLC chromatograms. (b) EPR spectra of adducts of DEPMPO spin trap (5 mM), illustrating the capacity of Tris to remove hydroxyl radical (HO* is produced in the Fenton reaction: Fe²⁺ (0.4 mM) + H₂O₂ (1.2 mM)). Spectrum in 10 mM Tris is composed of DEPMPO adducts with HO* (65%) and Tris-derived C-centred radical (35%), as determined by spectral simulation (dashed line). No EPR signal could be observed in 100 mM Tris. (c) The concentration of Epi following 5 min incubation with different [Fe²⁺]_i in 100 mM Tris buffer. Inset: Epi peaks in HPLC chromatograms.(d) UV-Vis spectra of Epi/Fe²⁺ systems after 5 min incubation in 100 mM Tris. No further changes were observed. Dashed line represents the subtraction of experimental spectra. The resulting spectrum with $\lambda_{max} = 545$ nm in the system with [Epi]/[Fe²⁺]_i = 0.5, represents the sum of the spectrum for [Epi]/[Fe²⁺]_i = 1 and the spectrum of [Fe³⁺] = 0.2 mM.

Figure S6 Schematic presentation of the chemical structures of Epi–Fe³⁺ complexes. L – other ligands (e.g. OH⁻, HPO₄²⁻, H₂PO₄⁻, H₂O).

Table S1 Reactions that are relevant for Fe²⁺ oxidation at pH 7.4.

No	Reaction	k (M ⁻¹ s ⁻¹)	Ref
1	$Fe^{2+} + O_2 \rightarrow Fe^{3+} + O_2^{-+}$	4 × 10 ⁻²	(a)
2	$Fe^{2+} + O_2^{-+} + 2H^+ \rightarrow Fe^{3+} + H_2O_2$	1 × 10 ⁷	
3	$Fe^{3+} + O_2^{-+} \rightarrow Fe^{2+} + O_2$	1.5×10^8	
4	$Fe^{2+} + H_2O_2 \rightarrow Fe^{3+} + HO^{\bullet} + OH^{-}$	10 ²	(b)
5	$Fe^{2+} + HO^{\bullet} \rightarrow FeOH^{2+}$	3.2×10^8	
6**	$2H_2O_2 (CAT) \rightarrow 2H_2O + O_2$		

Fenton reaction; The mechanism of catalase-mediated degradation of H_2O_2 . The concentration of accumulated H_2O_2 is calculated as $2\times\Delta[O_2]$ that is induced by CAT.

- (a) King, D. W.; Lounsbury, H. A.; Millero, F. J. Environ. Sci. Technol., 1995, 29, 818-825.
- (b) Halliwell, B. & Gutterridge, J. M. C. Free Radicals in Biology and Medicine, 4th ed, Clarendon Press, Oxford, 2007.