

Supplementary material for the article:

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Supporting information

Structural differences in diarylheptanoids analogues from *Alnus viridis* and *Alnus glutinosa* influence their activity and selectivity towards cancer cells

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Spectroscopic data for new compound v4 isolated from *A. viridis*.

(3S)-1,7-di(4-hydroxyphenyl)-heptan-3-*O*- α -L-arabinofuranosyl(1→6)- β -D-glucopyranoside (**v4**): brown amorphous solid; $[\alpha]_D^{22}$ -51.0 (c 0.1, MeOH); UV (MeOH) λ_{max} (log ε) 224 (3.83), 278 (3.55) nm; IR (MeOH) ν_{max} 3357, 2929, 1614, 1513, 1450, 1361, 1233, 1076, 1042 cm⁻¹; ¹H-NMR and ¹³C-NMR data, see Table S1; HR-ESI-MS *m/z* 593.2617 [M - H]⁻ (calcd. for C₃₀H₄₂O₁₂-H, 593.2604).

Experimental procedure for isolation and structure elucidation. Optical rotations were measured on a Rudolph Research Analytical AUTOPOL IV automatic polarimeter. UV spectra were recorded using GBC Cintra 40 UV/VIS spectrometer. IR spectra were recorded at a ThermoScientific Nicolet 6700 FT-IR spectrometer using capilar film technique. All NMR spectra (¹H, ¹³C, HSQC, HMBC) were recorded on a Bruker Avance III 500 spectrometer at 500.26 for ¹H and 125.80 MHz for ¹³C, with CD₃OD as solvent and TMS as reference. Mass spectral (HR-ESI-MS) data were obtained from Agilent Technologies 6210 time-of-flight LC/MS system. For column chromatography (CC) silicagel 60 (SiO₂; under 0.063 mm, Merck) was used. Analytical TLC was carried out on silicagel 60 GF₂₅₄ 20 × 20 cm plates, layer thickness 0.25 mm (Merck). Semipreparative HPLC was performed on an Agilent 1100 Series instrument equipped with DAD and using a Zorbax Eclipse XDB C-18 column (250 mm × 9.4 mm, 5 μm). All solvents used for column chromatography were freshly distilled and solvents for HPLC analysis were chromatographic grade.

Plant Material. The plant material was collected at Stara planina in south-eastern Serbia in July 2010 and was identified by Professor Petar Marin, Faculty of Biology, University of Belgrade, Serbia. A voucher specimen No. 16681 was deposited at the Herbarium of the Institute of Botany and Botanical Garden 'Jevremovac', Faculty of Biology, University of Belgrade (BEOU), Serbia.

Extraction and isolation. The air-dried bark (150.0 g) was powdered and extracted with CHCl₃/MeOH 1:1 (4 × 1 l, 24 h) at room temperature, with the use of an ultrasonic bath in the last hour of extraction. The yield of the extraction was 22 %. The half of the crude extract (16.5 g) was fractionated by gradient CC (60 cm length and 5 cm diameter column size), starting elution with 100% CH₂Cl₂ and gradually increasing

polarity by addition of MeOH (up to 40% MeOH) to yield 200 fractions of approximately 15 ml. Similar fractions were combined after TLC (carried out with different CH₂Cl₂/MeOH solvent systems) and further fractionated by semipreparative HPLC-DAD into pure compounds using 0.025% HCOOH/MeCN elution system with flow rate of 4 ml/min and gradient program: 0-20 min, 15-25% MeCN; 20-25 min, 25-40% MeCN; 25-28 min, 40-70% MeCN; 28-31 min, 70-100%. The detection wavelength was 280 nm.

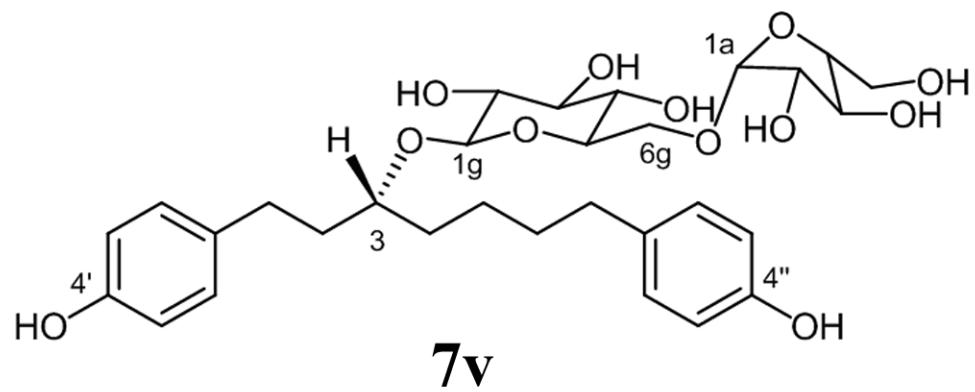
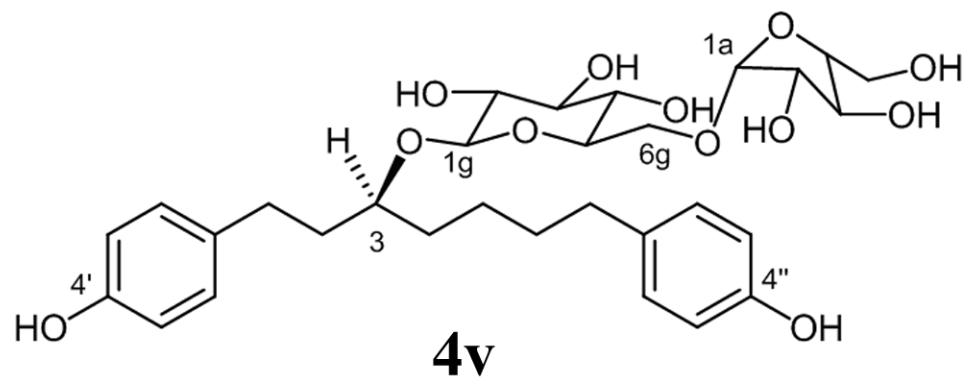


Figure S1. Chemical structures of compounds **4v** and **7v** from *A. viridis*.

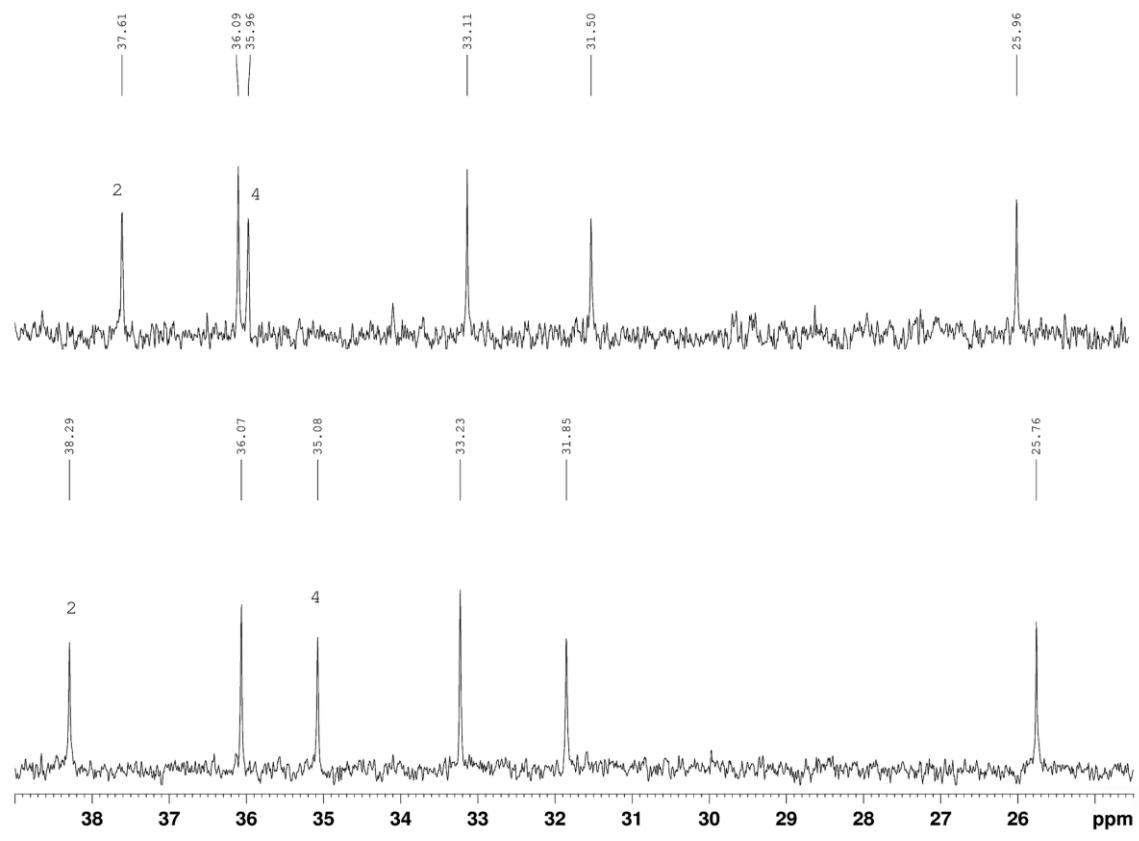


Figure S2. Heptane parts of the ^{13}C NMR spectra (CD_3OD) of **4v** (up) and **7v** (down).

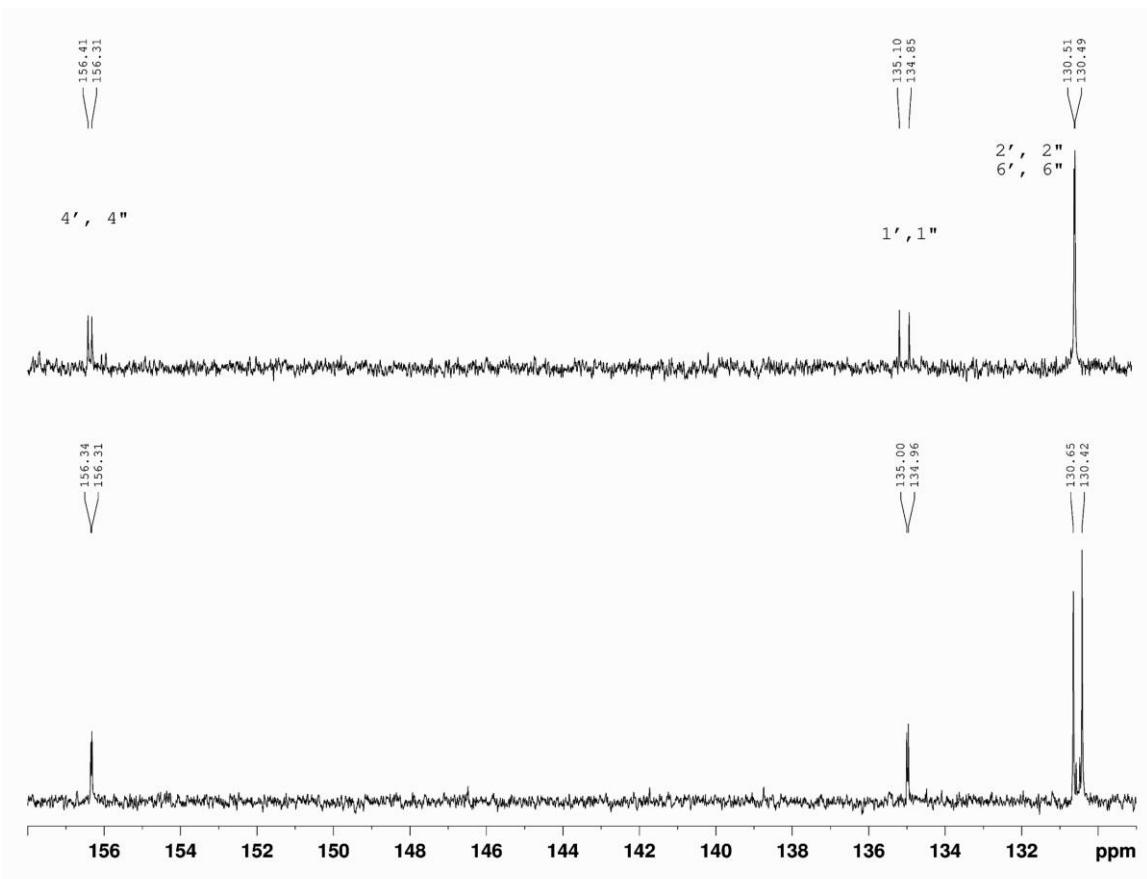


Figure S3. Aromatic parts of the ^{13}C NMR spectra (CD_3OD) of **4v** (up) and **7v** (down).

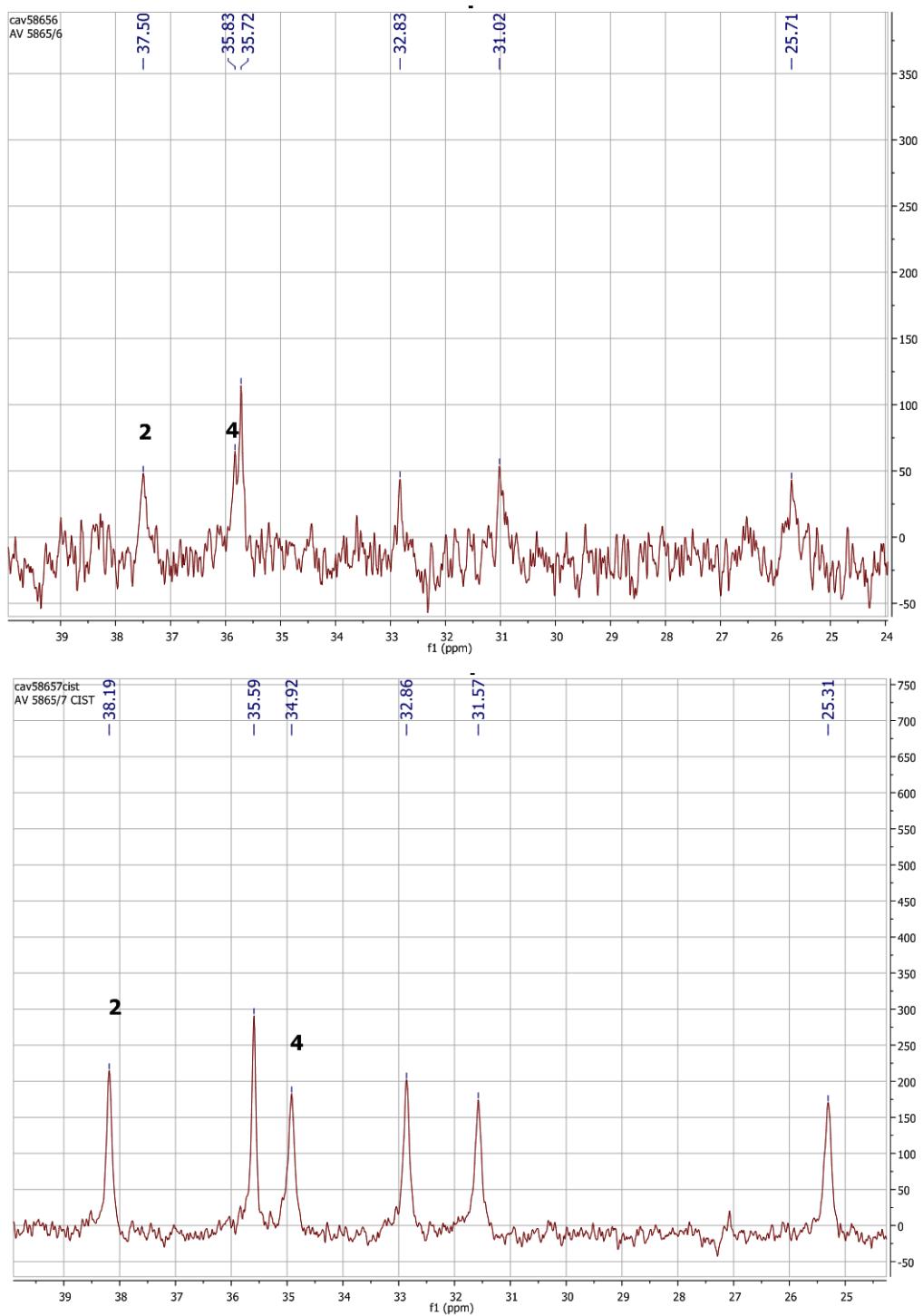


Figure S4. Heptane parts of the ^{13}C NMR spectra (pyridine-d5) of **4v** (up) and **7v** (down) used for comparing with (-)-centrolobol literature data.

Table S1. ^{13}C and ^1H NMR data (CD₃OD) for **4v** and **7v**.

C/H	4v		7v	
	δ_{C} , tip	δ_{H} (J u Hz)	δ_{C} , tip	δ_{H} (J u Hz)
1	31,5; CH ₂	2,61 m	31,9; CH ₂	2,60 m
2	37,6; CH ₂	1,77 m	38,3; CH ₂	1,75 m
3	80,4; CH	3,67 t (6,0)	80,2; CH	3,66 m
4	36,0; CH ₂	1,59 m	35,1; CH ₂	1,55 m; 1,61 m
5	26,0; CH ₂	1,40 m	25,8; CH ₂	1,39 m
6	33,1; CH ₂	1,59 m	33,2; CH ₂	1,54 m
7	36,1; CH ₂	2,51 t (7,0)	36,1; CH ₂	2,50 t (7,5)
1'	135,1; C	-	135,0; C	-
2'	130,5; CH	7,02 d (8,5)	130,6; CH	7,02 d (8,5)
3'	116,2; CH	6,68 d (8,5)	116,2; CH	6,69 d (8,5)
4'	156,4; C	-	156,3; C	-
5'	116,2; CH	6,68 d (8,5)	116,2; CH	6,69 d (8,5)
6'	130,5; CH	7,01 d (8,5)	130,6; CH	7,02 d (8,5)
1"	134,9; C	-	135,0; C	-
2"	130,5; CH	7,01 d (8,5)	130,4; CH	6,97 d (8,5)
3"	116,2; CH	6,68 d (8,5)	116,2; CH	6,68 d (8,5)
4"	156,3; C	-	156,3; C	-
5"	116,2; CH	6,68 d (8,5)	116,2; CH	6,68 d (8,5)
6"	130,5; CH	6,99 d (8,5)	130,4; CH	6,97 d (8,5)
GlcP				
1g	103,9; CH	4,29 d (8,0)	103,7; CH	4,29 d (8,0)
2g	75,5; CH	3,18 dd (8,0; 1,5)	75,4; CH	3,18 dd (8,0; 1,5)
3g	78,3; CH	3,34 m	78,2; CH	3,34 m
4g	72,3; CH	3,34 m	72,1; CH	3,34 m
5g	76,6; CH	3,36 m	76,6; CH	3,39 m
6g	68,3; CH ₂	3,61 m 3,99 m	68,2; CH ₂	3,64 dd (11,0; 5,5) 4,03 dd (11,0; 2,0)
Arabf				
1a	110,0; CH	4,96 d (1,0)	110,0; CH	4,97 d (1,0)
2a	83,2; CH	4,00 m	83,2; CH	4,01 dd (3,5; 1,5)
3a	79,1; CH	3,84 m	79,0; CH	3,84 dd (5,5; 3,5)
4a	86,2; CH	3,96 m	86,1; CH	3,96 m
5a	63,1; CH ₂	3,58 m 3,72 dd (12,0; 3,5)	63,0; CH ₂	3,59 dd (12,0; 5,5) 3,69 dd (12,0; 3,5)

GlcP – glucopyranosyl group; **Arabf** – arabinofuranosyl group

Table S2. Statistical analysis of growth inhibition activity of *A. viridis* and *A. glutinosa* diarylheptanoids in NCI-H460 and HaCaT cells.

NCI-H460													
	1v	2v	3v	4v	5v	6v	7v	8v	9v	3g	5g	8g	9g
1v	/	****	****	****	****	****	****	****	****	****	****	****	****
2v	****	/	****	****	****	****	*	****	****	****	****	****	****
3v	****	****	/	****	ns	****	****	ns	ns	ns	ns	ns	ns
4v	****	****	****	/	****	****	****	****	****	****	****	****	****
5v	****	****	ns	****	/	****	****	*	ns	ns	*	ns	ns
6v	****	****	****	****	****	/	****	****	****	****	****	****	****
7v	****	*	****	****	****	****	/	****	****	****	****	****	****
8v	****	****	ns	****	*	****	****	/	****	ns	ns	ns	ns
9v	****	****	ns	****	ns	****	****	****	/	ns	***	**	ns
3g	****	****	ns	****	ns	****	****	ns	ns	/	ns	ns	ns
5g	****	****	ns	****	*	****	****	ns	***	ns	/	ns	ns
8g	****	****	ns	****	ns	****	****	ns	**	ns	ns	/	ns
9g	****	****	ns	****	ns	****	****	ns	ns	ns	ns	ns	/
HaCaT													
	1v	2v	3v	4v	5v	6v	7v	8v	9v	3g	5g	8g	9g
1v	/	ns	****	ns	****	****	****	****	****	****	***	****	****
2v	ns	/	****	ns	****	ns	**	****	****	****	ns	****	****
3v	****	****	/	****	ns	****	****	****	ns	****	****	ns	****
4v	ns	ns	****	/	****	**	***	****	****	****	ns	****	****
5v	****	****	ns	****	/	****	****	****	ns	***	****	ns	****
6v	****	ns	****	**	****	/	ns	****	****	****	ns	****	****
7v	****	**	****	***	****	ns	/	****	****	****	ns	****	****
8v	****	****	****	****	****	****	****	/	****	ns	****	****	ns
9v	****	****	ns	****	ns	****	****	****	/	***	****	ns	****
3g	****	****	****	****	***	****	****	ns	***	/	****	***	ns
5g	***	ns	****	ns	****	ns	ns	****	****	****	/	****	****
8g	****	****	ns	****	ns	****	****	****	ns	***	****	/	****
9g	****	****	****	****	****	****	****	ns	****	ns	****	****	/

* p<0.05

** p<0.01

*** p<0.001

**** p<0.0001

ns = non significant

Table S3. Statistical analysis of cell death inducing activity of *A. viridis* and *A. glutinosa* diarylheptanoids in NCI-H460 and HaCaT cells.

NCI-H460									
		5v 15 µM	5v 45 µM	5g 15 µM	5g 45 µM	9v 15 µM	9v 45 µM	9g 15 µM	9g 45 µM
viable cells	untreated	ns	****	ns	****	*	****	ns	****
early apoptosis		ns	**	ns	ns	ns	ns	ns	***
late apoptosis		ns	ns	ns	*	ns	****	ns	**
necrosis		ns	ns	ns	*	ns	ns	ns	*
HaCaT									
		5v 15 µM	5v 45 µM	5g 15 µM	5g 45 µM	9v 15 µM	9v 45 µM	9g 15 µM	9g 45 µM
viable cells	untreated	***	****	ns	**	****	****	ns	****
early apoptosis		ns							
late apoptosis		ns	****	ns	ns	*	****	ns	****
necrosis		ns	****	ns	ns	ns	*	ns	***

* p<0.05

** p<0.01

*** p<0.001

**** p<0.0001

ns = non significant

Table S4. Statistical analysis of cell cycle arresting activity of *A. viridis* and *A. glutinosa* diarylheptanoids in NCI-H460 and HaCaT cells.

NCI-H460									
		5v 15 µM	5v 45 µM	5g 15 µM	5g 45 µM	9v 15 µM	9v 45 µM	9g 15 µM	9g 45 µM
subG ₀	untreated	ns	**	ns	ns	*	*	ns	ns
G ₀ /G ₁		ns	****	ns	ns	***	****	ns	ns
S		ns							
G ₂ /M		ns							
HaCaT									
		5v 15 µM	5v 45 µM	5g 15 µM	5g 45 µM	9v 15 µM	9v 45 µM	9g 15 µM	9g 45 µM
subG ₀	untreated	*	*	ns	ns	ns	ns	ns	ns
G ₀ /G ₁		ns	**	ns	ns	**	*	ns	**
S		ns							
G ₂ /M		*	ns						

* p<0.05

** p<0.01

*** p<0.001

**** p<0.0001

ns = non significant