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## Ruthenium-arene complexes with NSAIDs: Synthesis, characterization and bioactivity

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## Supplementary material

## S1. NMR spectra of synthesized complexes




Figure S1. Parallel ${ }^{1} \mathrm{H}$ NMR spectra of ligand1 and complex 1


Figure S2. Parallel ${ }^{13} \mathrm{C}$ NMR spectra of ligand 1 and complex 1


Figure S3. Parallel ${ }^{1} \mathrm{H}$ NMR spectra of ligand 2 and complex 2
complex 2
ligand 2



Figure S4. Parallel ${ }^{13} \mathrm{C}$ NMR spectra of ligand 2 and complex 2



Figure S5. Parallel ${ }^{1} \mathrm{H}$ NMR spectra of ligand1 and complex 3


Figure S6. Parallel ${ }^{13} \mathrm{C}$ NMR spectra of ligand 1 and complex 3


Figure S7. Parallel ${ }^{1} \mathrm{H}$ NMR spectra of ligand 2 and complex 4
complex 4


ligand 2


Figure S8. Parallel ${ }^{13} \mathrm{C}$ NMR spectra of ligand 2 and complex 4

## S2. MTT assay

Table 1. $\mathrm{IC}_{50}[\mu \mathrm{M}]$ values obtained after 72 h of continuous drug action.

| Compound | K562 | A549 | MDA-MB-231 | MRC-5 | *SI $_{\text {K562 }}$ | *SI $_{\text {A549 }}$ | *SI $_{\text {MDA-MB-231 }}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\mathbf{1}$ | $11.9 \pm 4.4$ | $45.5 \pm 2.7$ | $22 \pm 3.6$ | $39.6 \pm 3.7$ | 3.33 | 0.87 | 1.80 |
| $\mathbf{2}$ | $96.4 \pm 2$ | $145.1 \pm 6.4$ | $153 \pm 1.2$ | $222.6 \pm 23.9$ | 2.31 | 1.53 | 1.45 |
| $\mathbf{3}$ | $13.2 \pm 6.2$ | $31.7 \pm 1.15$ | $26 \pm 1.7$ | $42 \pm 1.3$ | 3.18 | 1.32 | 1.62 |
| $\mathbf{4}$ | $133 \pm 7$ | $142.4 \pm 9.3$ | $121.4 \pm 1.8$ | $275.7 \pm 14.5$ | 2.07 | 1.94 | 2.27 |
| Hindo | $155.9 \pm 11.4$ | $161.5 \pm 13.9$ | $244.7 \pm 17.8$ | $230.5 \pm 17.8$ | 1.48 | 1.43 | 0.94 |
| Hmef | $143.9 \pm 4.1$ | $217.3 \pm 46.7$ | $237.9 \pm 18.8$ | $>300$ | $>2.08$ | $>1.38$ | $>1.26$ |
| CDDP | $10.3 \pm 1.2$ | $13.6 \pm 1.8$ | $15.9 \pm 2.1$ | $9.3 \pm 0.9$ | 0.90 | 0.68 | 0.58 |

* $\mathrm{IC}_{50}[\mu \mathrm{M}]$ values are presented as the mean $\pm$ SEM of three independent experiments. > 300 denotes that $\mathrm{IC}_{50}$ was not obtained in the range of concentrations tested up to $300 \mu \mathrm{M}$.
*SI-selectivity index for tested complexes, ligands and cisplatin, in tumor cell lines (K562, A549 and MDA-MB-231), related to non-tumor MRC-5 cell line: $\mathrm{SI}_{\mathrm{K} 562}\left(\mathrm{IC}_{50} \mathrm{MRC}-5 / \mathrm{IC}_{50} \mathrm{~K}_{562}\right), \mathrm{SI}_{\mathrm{A}_{549}}$ ( $\mathrm{IC}_{50}$ MRC-5/IC $\mathrm{S}_{50}$ A549), $\mathrm{SI}_{\mathrm{MDA}}$-MB-231 $\left(\mathrm{IC}_{50} \mathrm{MRC}^{2} / \mathrm{IC}_{50}\right.$ MDA-MB-231). SI values for tested complexes and ligands were obviously higher than for cisplatin, particularly in MDA-MB-231 and K562.


## S3. Interaction with biomolecules

## S3-1. Interaction with serum albumins

The extent of the inner-filter effect can be roughly estimated with the following formula:

$$
\begin{equation*}
I_{\text {corr }}=I_{\text {meas }} \times 10^{\frac{\varepsilon\left(\lambda_{\text {ex }}\right) \mathrm{cd}}{2}} \times 10^{\frac{\varepsilon\left(\lambda_{\text {em }}\right) \mathrm{cd}}{2}} \tag{eq.S1}
\end{equation*}
$$

where $\mathrm{I}_{\text {corr }}=$ corrected intensity, $\mathrm{I}_{\text {meas }}=$ the measured intensity, $\mathrm{c}=$ the concentration of the quencher, $\mathrm{d}=$ the cuvette $(1 \mathrm{~cm}), \varepsilon\left(\lambda_{\text {exc }}\right)$ and $\varepsilon\left(\lambda_{\mathrm{em}}\right)=$ the $\varepsilon$ of the quencher at the excitation and the emission wavelength, respectively, as calculated from the UV-vis spectra of the complexes. ${ }^{1}$

The Stern-Volmer and Scatchard graphs are used in order to study the interaction of a quencher with serum albumins. According to Stern-Volmer quenching equation: ${ }^{2}$

$$
\begin{equation*}
\frac{\mathrm{I}}{\mathrm{I}}=1+\mathrm{k}_{\mathrm{q}} \tau_{0}[\mathrm{Q}]=1+\mathrm{K}_{\mathrm{sv}}[\mathrm{Q}] \tag{eq.S2}
\end{equation*}
$$

where $\mathrm{Io}=$ the initial tryptophan fluorescence intensity of SA, $\mathrm{I}=$ the tryptophan fluorescence intensity of SA after the addition of the quencher (i.e. complexes $\mathbf{1 - 4}$ ), $\mathrm{k}_{\mathrm{q}}=$ the quenching constant, $\mathrm{K}_{\mathrm{SV}}=$ the Stern-Volmer constant, $\tau_{\mathrm{o}}=$ the average lifetime of SA without the quencher, $[\mathrm{Q}]=$ the concentration of the quencher) $\mathrm{K}_{\mathrm{SV}}$ (in $\mathrm{M}^{-1}$ ) can be obtained by the slope of the diagram Io/I versus $[\mathrm{Q}]$, and subsequently the quenching constant $\left(\mathrm{k}_{\mathrm{q}}\right.$, in $\left.\mathrm{M}^{-1} \mathrm{~s}^{-1}\right)$ is calculated from eq. S 3 , with $\tau_{\mathrm{o}}=10^{-8}$ s as fluorescence lifetime of tryptophan in SA,

$$
\begin{equation*}
\mathrm{K}_{\mathrm{SV}}=\mathrm{k}_{\mathrm{q}} \tau_{\mathrm{o}} \tag{eq.S3}
\end{equation*}
$$

From the Scatchard equation: ${ }^{3}$

$$
\begin{equation*}
\frac{\Delta \mathrm{I} / \mathrm{Io}}{[\mathrm{Q}]}=\mathrm{nK}-\mathrm{K} \frac{\Delta \mathrm{I}}{\mathrm{Io}} \tag{eq.S4}
\end{equation*}
$$

where n is the number of binding sites per albumin and K is the SA-binding constant, K (in $\mathrm{M}^{-1}$ ) is calculated from the slope in plots $(\Delta \mathrm{I} / \mathrm{Io}) /[\mathrm{Q}]$ versus $\Delta \mathrm{I} / \mathrm{Io}$ and n is given by the ratio of y intercept to the slope. ${ }^{3}$

## S3-2. Interaction with CT DNA

The DNA-binding constant $\left(\mathrm{K}_{\mathrm{b}}\right.$, in $\left.\mathrm{M}^{-1}\right)$ can be obtained by monitoring the changes in the absorbance at the corresponding $\lambda_{\max }$ with increasing concentrations of CT DNA and it is given by the ratio of slope to the $y$ intercept in plots $[\mathrm{DNA}] /\left(\varepsilon_{A}-\varepsilon_{f}\right)$ versus [DNA], according to the WolfeShimer equation: ${ }^{4}$

$$
\begin{equation*}
\frac{[\mathrm{DNA}]}{\left(\varepsilon_{\mathrm{A}}-\varepsilon_{\mathrm{f}}\right)}=\frac{[\mathrm{DNA}]}{\left(\varepsilon_{\mathrm{b}}-\varepsilon_{\mathrm{f}}\right)}+\frac{1}{\mathrm{~K}_{\mathrm{b}}\left(\varepsilon_{\mathrm{b}}-\varepsilon_{\mathrm{f}}\right)} \tag{eq.S5}
\end{equation*}
$$

where [DNA] is the concentration of DNA in base pairs, $\varepsilon_{\mathrm{A}}=\mathrm{A}_{\mathrm{obsd}} /[$ compound $], \varepsilon_{\mathrm{f}}=$ the extinction coefficient for the free compound and $\varepsilon_{b}=$ the extinction coefficient for the compound in the fully bound form.

## S3-3. Competitive studies with EB

The Stern-Volmer constant ( $\mathrm{K}_{\mathrm{sv}}$, in $\mathrm{M}^{-1}$ ) is used to evaluate the quenching efficiency for each compound according to the Stern-Volmer equation (eq. S2), ${ }^{2}$ where Io and I are the emission intensities of the EB-DNA solution in the absence and the presence of the quencher, respectively, [Q] is the concentration of the quencher (i.e. complexes 1-4), $\tau_{0}=$ the average lifetime of the emitting system without the quencher and $\mathrm{k}_{\mathrm{q}}=$ the quenching constant. $\mathrm{K}_{\text {sv }}$ may be obtained from the Stern-Volmer plots by the slope of the diagram Io/I versus [Q]. Taking $\tau_{o}=23 \mathrm{~ns}$ as the fluorescence lifetime of the EB-DNA system, ${ }^{5}$ the quenching constants ( $\mathrm{k}_{\mathrm{q}}$, in $\mathrm{M}^{-1} \mathrm{~s}^{-1}$ ) of the compounds can be determined according to eq. (S3).

## References

1 L. Stella, A.L. Capodilupo and M. Bietti, Chem. Commun., 2008, 4744.
2 J.R. Lakowicz, Principles of Fluorescence Spectroscopy, third ed., Plenum Press, New York, 2006.
3 Y. Wang, H. Zhang, G. Zhang, W. Tao and S. Tang, J. Luminescence, 2007, 126, 211.
4 A. Wolfe, G. Shimer and T. Meehan, Biochemistry, 1987, 26, 6392.
5 D.P. Heller and C.L. Greenstock, Biophys. Chem., 1994, 50, 305.

Table S2. The BSA and HSA binding constants and parameters ( $\mathrm{K}_{\mathrm{sv}}, \mathrm{k}_{\mathrm{q}}, \mathrm{K}, \mathrm{n}$ ) for complexes 1-4.

| Compound | $\mathbf{K s v}\left(\mathbf{M}^{-1}\right)$ | $\mathbf{k}_{\mathbf{q}}\left(\mathbf{M}^{-1} \mathbf{s}^{-1}\right)$ | $\mathbf{K}\left(\mathbf{M}^{-1}\right)$ | $\mathbf{n}$ |
| :--- | :--- | :--- | :--- | :--- |
| BSA |  |  |  |  |
| $\mathrm{K}\left[\mathrm{Ru}\left(\eta^{6}-p\right.\right.$-cymene $)($ indo $\left.) \mathrm{Cl}_{2}\right], \mathbf{1}$ | $1.25( \pm 0.05) \times 10^{5}$ | $1.25( \pm 0.05) \times 10^{13}$ | $4.49( \pm 0.30) \times 10^{5}$ | 0.68 |
| $\left(\mathrm{NH}_{4}\right)\left[\mathrm{Ru}\left(\eta^{6}-p\right.\right.$-cymene $\left.)\left(\mathrm{mef}^{2}\right) \mathrm{Cl}_{2}\right], \mathbf{2}$ | $1.70( \pm 0.08) \times 10^{5}$ | $1.70( \pm 0.08) \times 10^{13}$ | $3.63( \pm 0.15) \times 10^{5}$ | 0.86 |
| $\mathrm{~K}\left[\mathrm{Ru}\left(\eta^{6}-p\right.\right.$-toluene $)($ indo $\left.) \mathrm{Cl}_{2}\right], \mathbf{3}$ | $4.85( \pm 0.11) \times 10^{4}$ | $4.85( \pm 0.11) \times 10^{12}$ | $5.30( \pm 0.18) \times 10^{4}$ | 0.96 |
| $\left(\mathrm{NH}_{4}\right)\left[\mathrm{Ru}\left(\eta^{6}-p\right.\right.$-toluene $\left.\left.)(\mathrm{mef})\right) \mathrm{Cl}_{2}\right], \mathbf{4}$ | $1.30( \pm 0.04) \times 10^{5}$ | $1.30( \pm 0.04) \times 10^{13}$ | $2.63( \pm 0.10) \times 10^{5}$ | 0.84 |
| HSA |  |  |  |  |
| $\mathrm{K}\left[\mathrm{Ru}\left(\eta^{6}-p\right.\right.$-cymene $)($ indo $\left.) \mathrm{Cl}_{2}\right], \mathbf{1}$ | $6.10( \pm 0.29) \times 10^{4}$ | $6.10( \pm 0.29) \times 10^{12}$ | $2.15( \pm 0.08) \times 10^{5}$ | 0.57 |
| $\left(\mathrm{NH}_{4}\right)\left[\mathrm{Ru}\left(\eta^{6}-p\right.\right.$-cymene $\left.)\left(\mathrm{mef}^{2}\right) \mathrm{Cl}_{2}\right], \mathbf{2}$ | $5.46( \pm 0.19) \times 10^{4}$ | $5.46( \pm 0.19) \times 10^{12}$ | $9.79( \pm 0.34) \times 10^{4}$ | 0.78 |
| $\mathrm{~K}\left[\mathrm{Ru}\left(\eta^{6}-p\right.\right.$-toluene $)($ indo $\left.) \mathrm{Cl}_{2}\right], \mathbf{3}$ | $2.04( \pm 0.13) \times 10^{4}$ | $2.04( \pm 0.13) \times 10^{12}$ | $9.44( \pm 0.40) \times 10^{4}$ | 0.31 |
| $\left(\mathrm{NH}_{4}\right)\left[\mathrm{Ru}\left(\eta^{6}-p\right.\right.$-toluene $\left.)\left(\mathrm{mef}^{2}\right) \mathrm{Cl}_{2}\right], \mathbf{4}$ | $4.32( \pm 0.29) \times 104$ | $4.32( \pm 0.29) \times 1012$ | $4.24( \pm 0.12) \times 10^{5}$ | 0.37 |



Figure S9. Stern-Volmer quenching plot of HSA for complexes (A)-(D) 1-4, respectively.


Figure S10. Stern-Volmer quenching plot of BSA for complexes (A)-(D) 1-4, respectively.


Figure S11. Scatchard plot of HSA for complexes (A)-(D) 1-4, respectively.


Figure S12. Scatchard plot of BSA for complexes (A)-(D) 1-4, respectively.


Figure S13. Plot of [DNA] $/\left(\varepsilon_{A}-\varepsilon_{f}\right)$ vs [DNA] for complexes (A)-(D) 1-4, respectively.


Figure S14. Stern-Volmer quenching plot of EB-DNA fluorescence for complexes (A)-(D) 1-4, respectively.

