

Supplementary data for article:

Filipović, N. R.; Bjelogrić, S. K.; Pelliccia, S.; Jovanović, V. B.; Kojić, M.; Senćanski, M.; La Regina, G.; Silvestri, R.; Muller, C. D.; Todorović, T. R. Selenotriapine - An Isostere of the Most Studied Thiosemicarbazone with Pronounced pro-Apoptotic Activity, Low Toxicity and Ability to Challenge Phenotype Reprogramming of 3-D Mammary Adenocarcinoma Tumors, 2017. <https://doi.org/10.1016/j.arabjc.2017.11.017>

SUPPLEMENTARY DATA

Selenotriapine – an isostere of the most studied thiosemicarbazone with pronounced pro-apoptotic activity, low toxicity and ability to challenge phenotype reprogramming of 3-D mammary adenocarcinoma tumors

Nenad R. Filipović^a, Snežana K. Bjelogrić^b, Sveva Pelliccia^c, Vesna B. Jovanović^d, Milan Kojić^e, Milan Senćanski^f, Giuseppe La Regina^g, Romano Silvestri^g, Christian D. Muller^{h,*}, Tamara R. Todorović^{d,*}

^a *Faculty of Agriculture, University of Belgrade, Nemanjina 6, Belgrade, Serbia*

^b *National Cancer Research Center of Serbia, Pasterova 14, Belgrade, Serbia*

^c *Dipartimento di Farmacia, Università di Napoli "Federico II", via D. Montesano 49, 80131 Naples, Italy*

^d *Faculty of Chemistry, University of Belgrade, Studentski trg 12-16, Belgrade, Serbia*

^e *Laboratory for Molecular Microbiology, Institute of Molecular Genetics and Genetic Engineering, University of Belgrade, Belgrade, Serbia*

^f *Center for Multidisciplinary Research, Institute of Nuclear Sciences "Vinča", University of Belgrade, Belgrade, Serbia*

^g *Dipartimento di Chimica e Tecnologie del Farmaco, Laboratory affiliated to Istituto Pasteur-Fondazione Cenci Bolognetti, Sapienza Università di Roma, Piazzale Aldo Moro 5, I-00185 Roma, Italy*

^h *Institut Pluridisciplinaire Hubert Curien, UMR 7178 CNRS Université de Strasbourg, 67401 Illkirch, France*

***Authors for correspondence:** Dr Tamara R. Todorović, Faculty of Chemistry, University of Belgrade, Studentskitrg 12-16, Belgrade, Serbia, +381(0)11-3336-731, tamarat@chem.bg.ac.rs

Dr Christian D. Muller, InstitutPluridisciplinaire Hubert Curien, UMR 7178 CNRS Université de Strasbourg, 67401 Illkirch, France, +33(0)6-88-27-57-39, cdmuller@unistra.fr

Content

Experimental	3
HSA binding experiments.....	3
Figure S1. UV/vis spectrum of Se-3-AP	4
Figure S2. Fluorescence spectrum of Se-3-AP	4
Figure S3. ¹ H-NMR spectrum of Se-3-AP	5
Figure S4. ¹³ C-NMR spectrum of Se-3-AP	5
Figure S5. COSY spectrum of Se-3-AP.....	6
Figure S6. ¹ H- ¹³ C HSQC spectrum of Se-3-AP.....	6
Figure S7. ¹ H- ¹³ C HMBC spectrum of Se-3-AP	7
Figure S8. ⁷⁷ Se-NMR spectrum of Se-3-AP.....	7
Figure S9. Cyclic voltammograms of Se-3-AP and 3-AP.....	8
Figure S10. Concentration-response curves	9
Figure S11. Changes in distribution of MCF-7 cells within phases of mitotic division.....	10
Figure S12. Stern-Volmer plot of F_0/F vs $[Q]$ at three different temperatures.....	11
Figure S13. Modified Stern-Volmer plot	11
Figure S14. The plot of $\ln K_b$ vs $1/T$ for the interaction of Se-3-AP with HSA	12
Figure S15. Double-log plot for determination of binding constants K_b , and the number of binding sites n at three temperatures.....	12
References	13

Experimental

HSA binding experiments. A ligand (quencher) can absorb energy at both the HSA excitation (280 nm) and emission (340 nm) wavelengths. In order to overcome the inner-filter effect, the absorbance values of the ligand used were measured and corresponding corrections were made during calculation of binding parameters according to the eq. (S1)^{S1}:

$$F_c = F_u \times 10^{(A_{ex} \times d_{ex} + A_{em} \times d_{em})/2} \quad (\text{eq. S1})$$

where F_u is the measured emission fluorescence intensity, F_c is the corrected fluorescence intensity that would be measured in the absence of any inner-filter effects, d_{ex} and d_{em} are the cell path lengths in the excitation and emission direction (1 cm), A_{ex} and A_{em} are the absorbance values of the quencher measured at the excitation and peak emission wavelength.

Fluorescence quenching data were processed using the Stern–Volmer eq. (S2):

$$F_0/F = 1 + K_{sv}[Q] = 1 + k_q\tau_0[Q] \quad (\text{eq. S2})$$

where F_0 and F are the HSA fluorescence intensities at 340 nm before and after addition of the quencher (Se-3-AP), K_{sv} is the Stern-Volmer quenching constant, k_q stands for the fluorescence quenching rate constant, τ_0 is the average fluorescence lifetime of the fluorophore (7.09 ns for HSA)^{S2} and $[Q]$ is the concentration of the quencher^{S3}. The quenching process was additionally analyzed using a modified Stern–Volmer eq. (S3)^{S3}:

$$F_0/F_0 - F = 1/f_a K_a [Q] + 1/f_a \quad (\text{eq. S3})$$

where F_0 and F are the HSA fluorescence intensities before and after addition of the quencher at concentration $[Q]$. K_a represents the effective quenching constant for the accessible fluorophores, and f_a is the fraction of accessible fluorophore.

Thermodynamic parameters of binding, the enthalpy (ΔH) and entropy change (ΔS), during the binding of Se-3-AP to HSA were determined by measuring the binding constants at three temperatures, and following the Van't Hoff eq. (S4):

$$\ln K_a = -\Delta H/RT + \Delta S/R \quad (\text{eq. S4})$$

where R is the universal gas constant, T is the temperature (in K), and K_a is the effective quenching constant at the corresponding temperature.

The estimations of association (binding) constants (K_b) and number of binding sites (n) of HSA and Se-3-AP were done using eq. (S5)^{S4}:

$$\log(F_0 - F)/F = -n \log(1/([Q] - [P]) \times (F_0 - F)/F_0) + n \log K_b \quad (\text{eq. S5})$$

where $[Q]$ and $[P]$ are total concentrations of ligand (Se-3-AP) and protein (HSA), respectively.

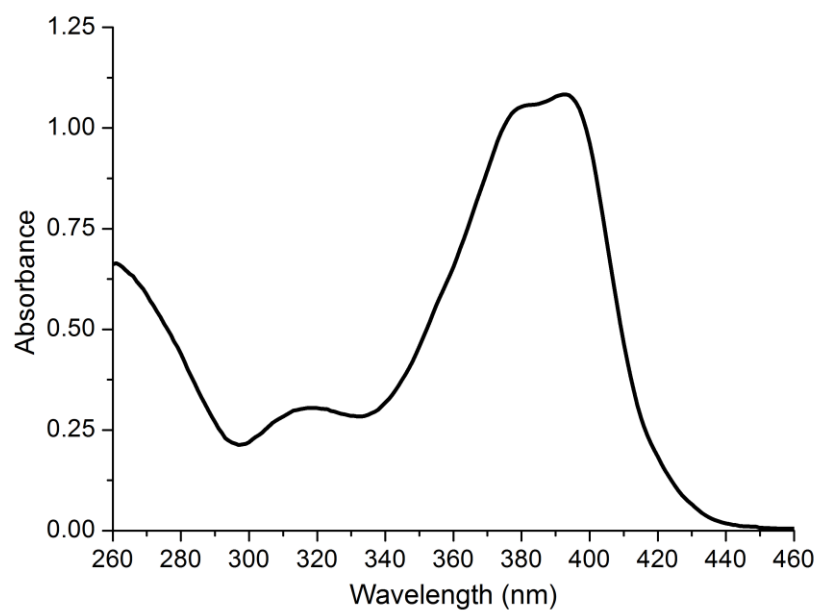


Figure S1. UV/vis spectrum of Se-3-AP (51 μ M) in DMSO.

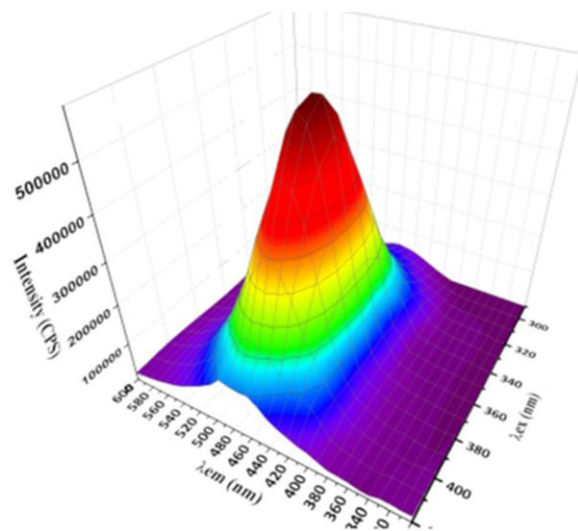


Figure S2. Fluorescence spectrum of Se-3-AP (10 mM, $\lambda_{\text{ex}} = 360$ nm) in DMSO.

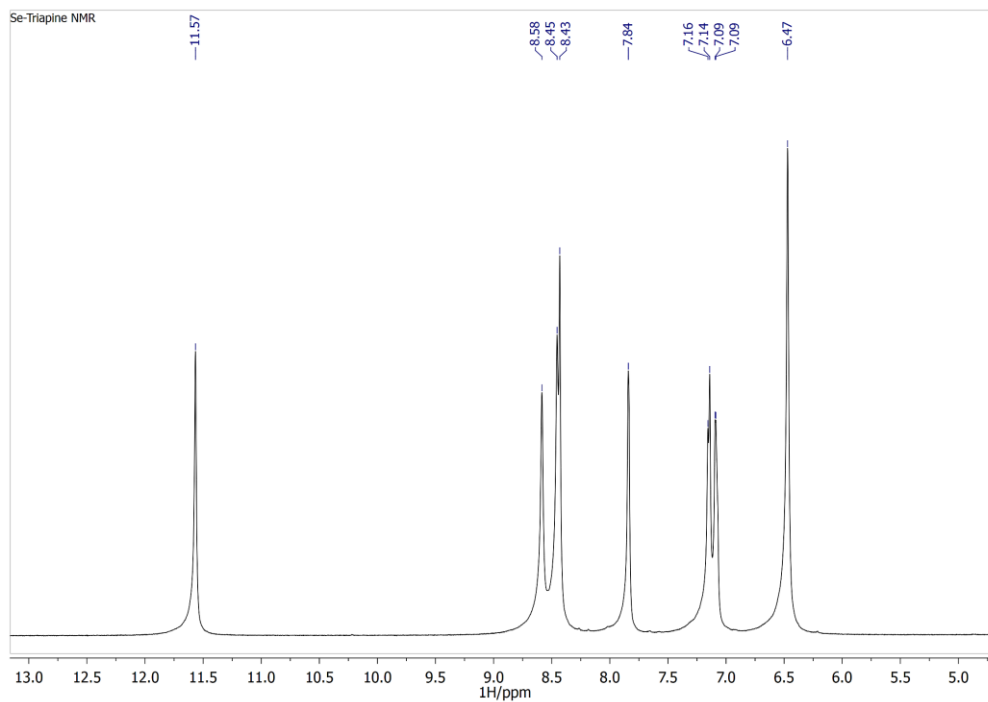


Figure S3. ^1H -NMR spectrum of Se-3-AP in $\text{DMSO-}d_6$.

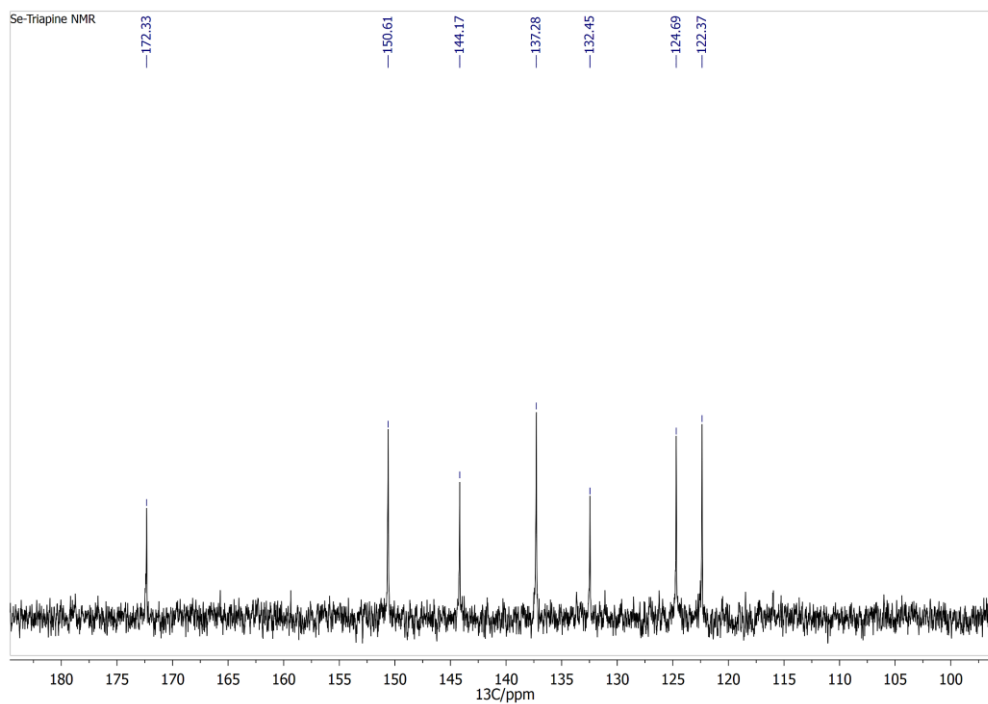


Figure S4. ^{13}C -NMR spectrum of Se-3-AP in $\text{DMSO-}d_6$.

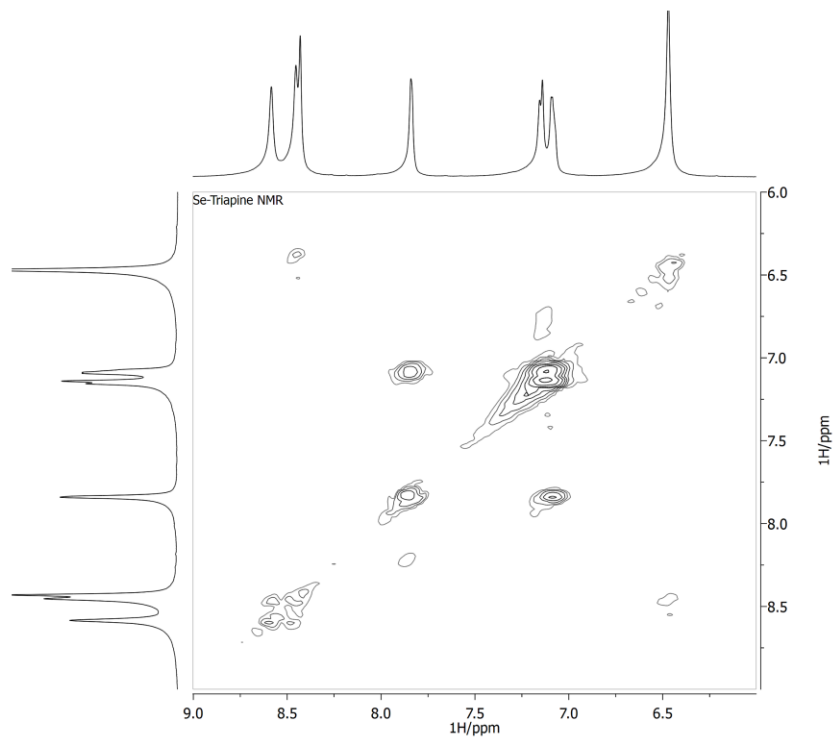


Figure S5. COSY spectrum of Se-3-AP in $\text{DMSO-}d_6$.

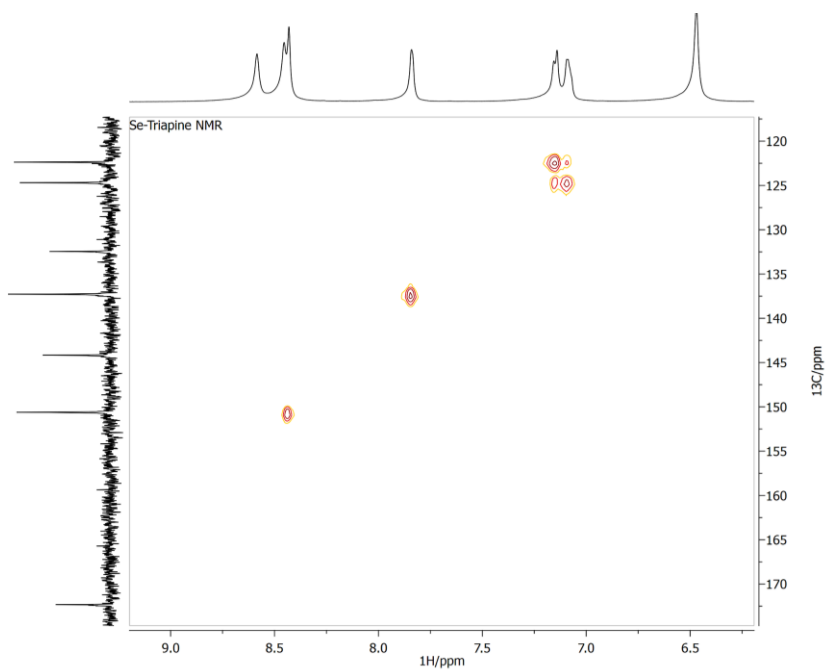


Figure S6. ^1H - ^{13}C HSQC spectrum of Se-3-AP in $\text{DMSO-}d_6$.

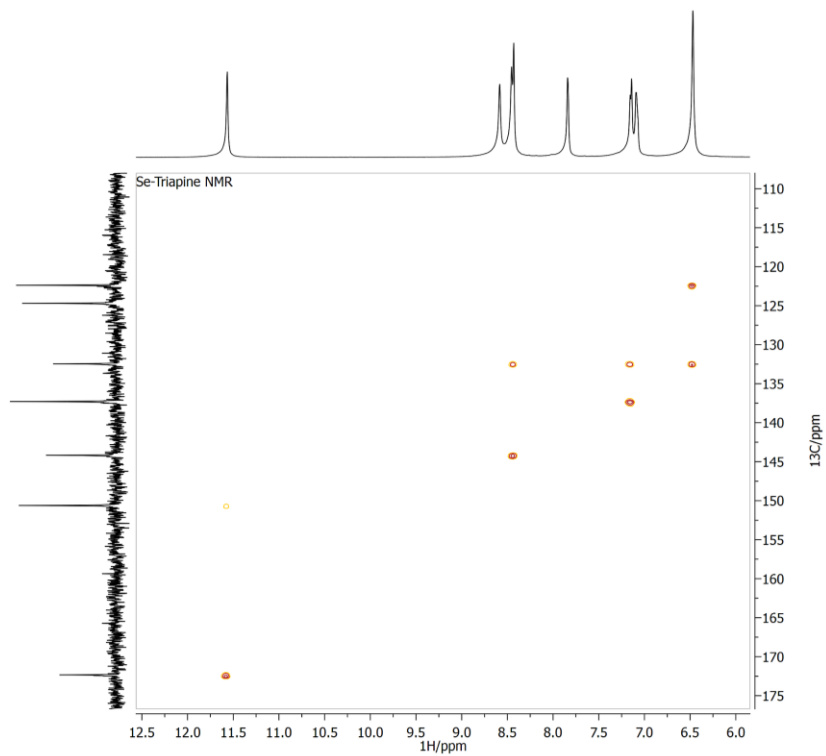


Figure S7. ^1H - ^{13}C HMBC spectrum of Se-3-AP in $\text{DMSO-}d_6$.

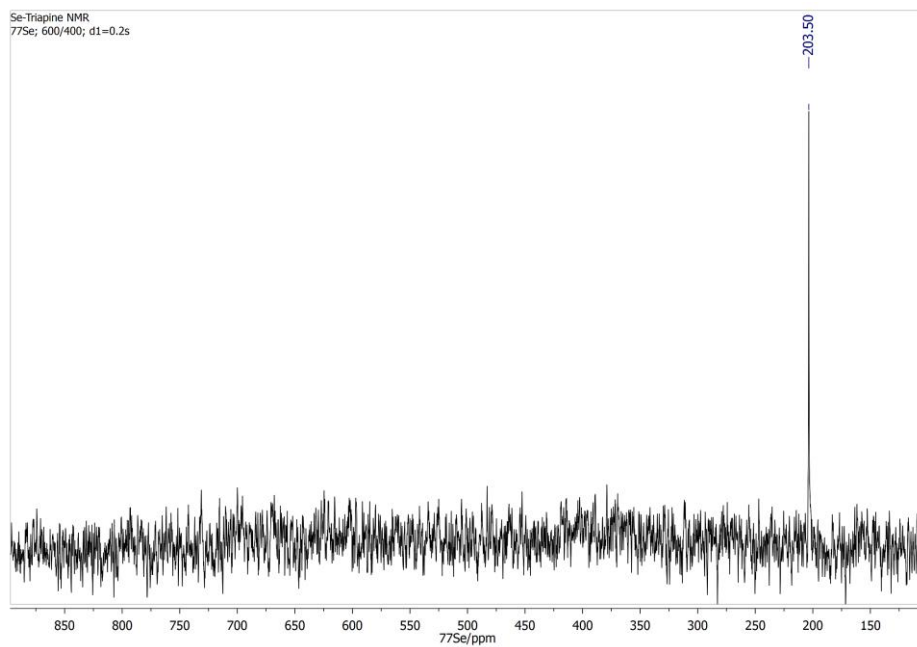


Figure S8. ^{77}Se -NMR spectrum of Se-3-AP in $\text{DMSO-}d_6$.

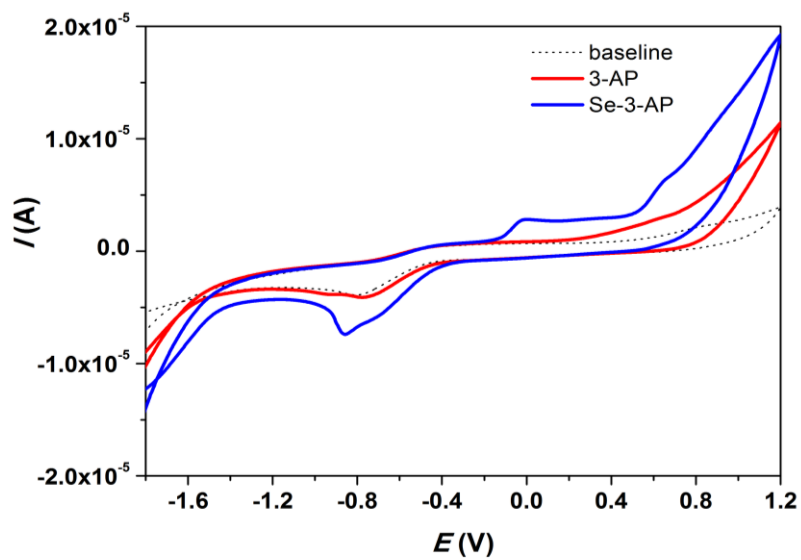


Figure S9. Cyclic voltammograms of Se-3-AP and 3-AP in 0.10 M $[n\text{-Bu}_4\text{N}]\text{PF}_6/\text{DMSO}$, GC electrode.

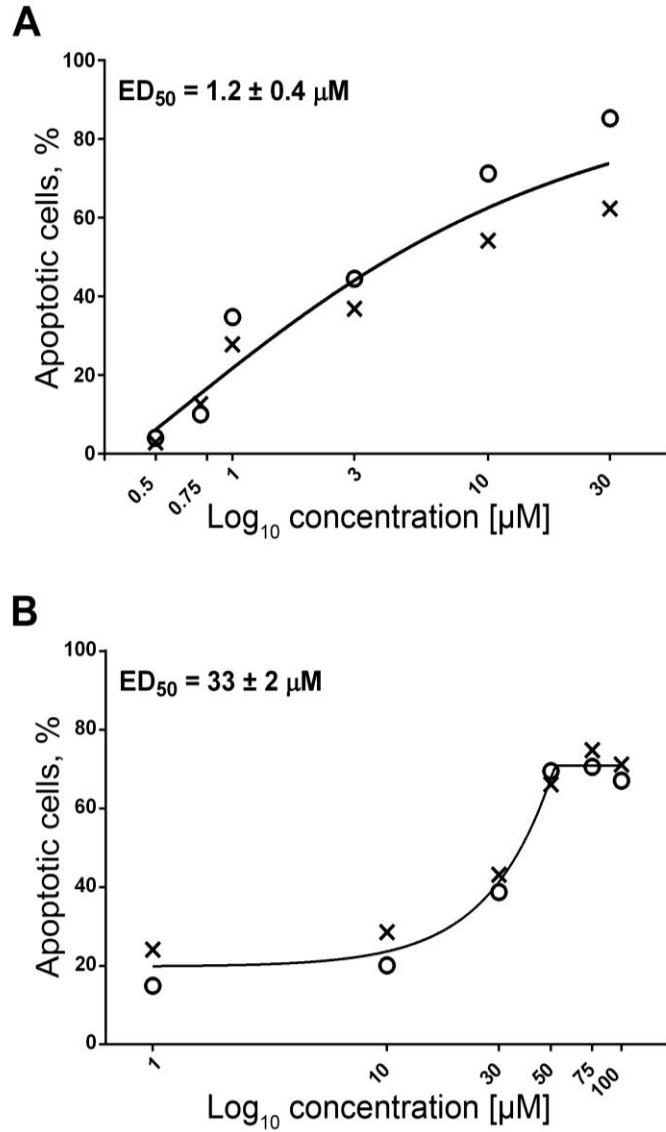


Figure S10. Concentration-response curves and ED₅₀ values for Se-3-AP on THP-1 (A) and MCF-7 cells (B) after 24 h treatment. Results are presented as percentages of apoptotic cells determined by means of Annexin V/propidium iodide double staining method for two independent experiments (circles and crosses), with asymmetric five-parameter sigmoidal curve computed for both replicates in GraphPad Prism software.

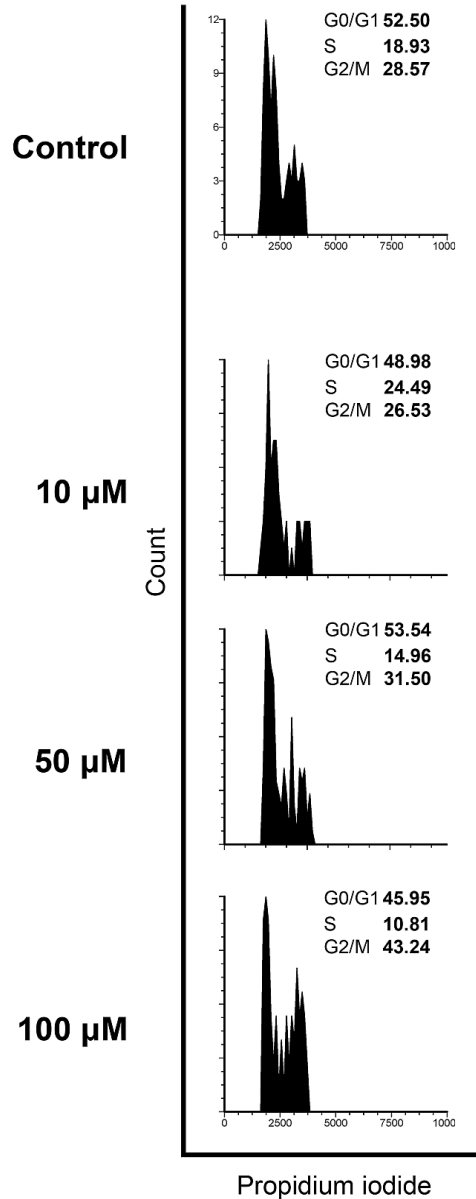


Figure S11. Changes in distribution of MCF-7 cells within phases of mitotic division induced by 24 h treatment with 3-AP. Analysis has been performed on the same cell samples represented in Figure 2A (right panel) on the remaining cells after Annexin V/PI read out. Incidences of cells found at the G0/G1, S and G2/M phases were determined according to non-treated control population. Results represent percentages of cells within phases of cell cycle obtained from a single experiment considering additional replicates have not been acquired due to 3-AP lack of activity.

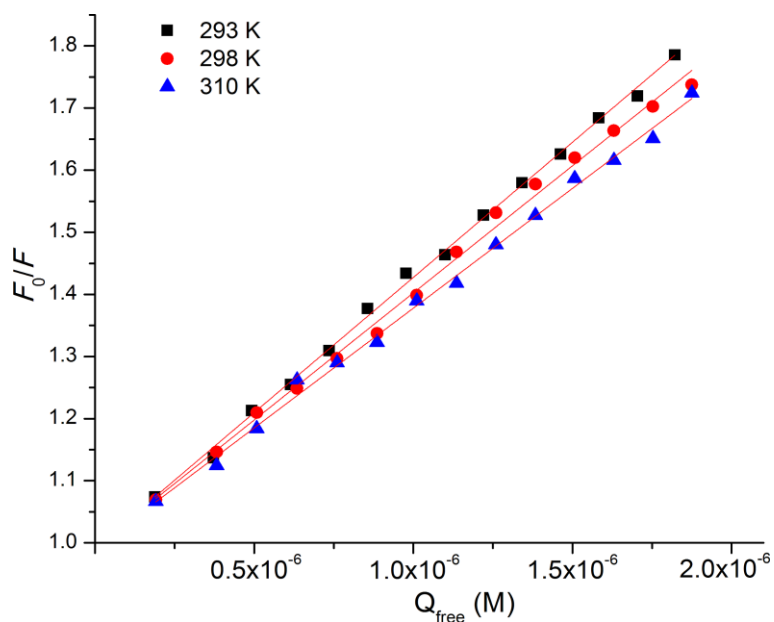


Figure S12. Stern-Volmer plot of F_0/F vs $[Q]$ at three different temperatures, where F_0 and F represent HSA fluorescence intensities in absence (F_0) and in the presence of the quencher (F), and $[Q]$ is the concentration of the quencher (Se-3-AP).

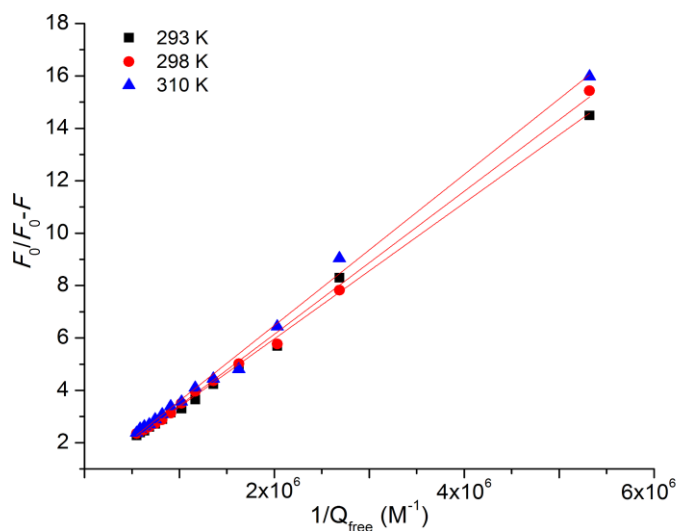


Figure S13. Modified Stern-Volmer plot for binding of Se-3-AP to HSA at three temperatures.

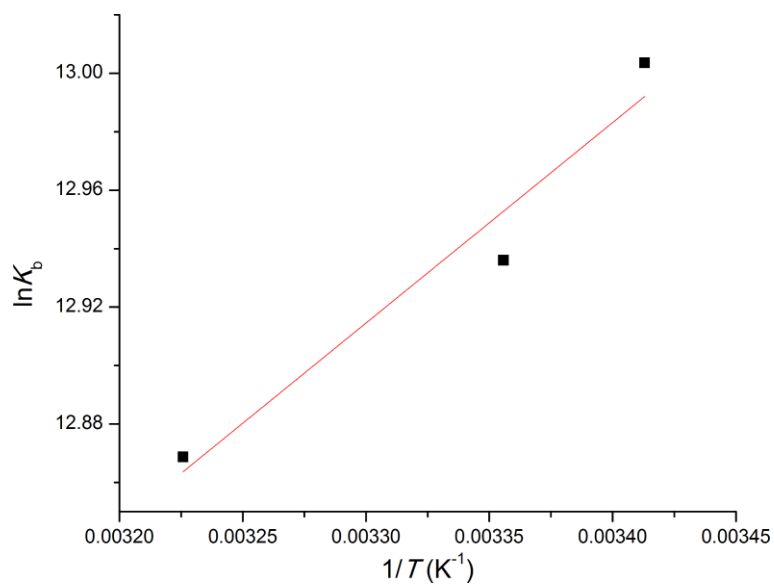


Figure S14. The plot of $\ln K_b$ vs $1/T$ for the interaction of Se-3-AP with HSA. K_b is given in M^{-1} .

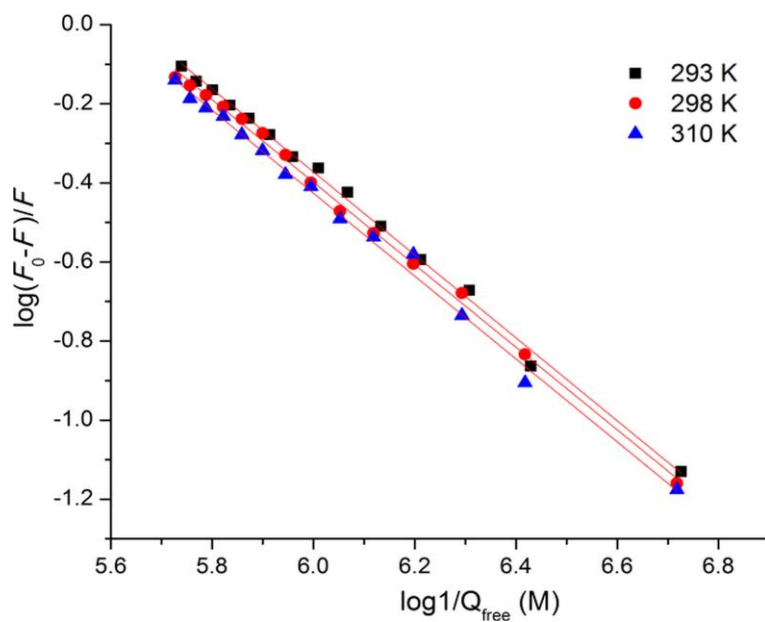


Figure S15. Double-log plot for determination of binding constants K_b , and the number of binding sites n at three temperatures; concentration of the quencher $[Q]$ is given in M .

References

- (S1) van de Weert, M.; Stella, L. Fluorescence Quenching and Ligand Binding: A Critical Discussion of a Popular Methodology. *J. Mol. Struct.***2011**, 998 (1), 144–150.
- (S2) Amiri, M.; Jankeje, K.; Albani, J. R. Origin of Fluorescence Lifetimes in Human Serum Albumin. Studies on Native and Denatured Protein. *J. Fluoresc.***2010**, 20 (3), 651–656.
- (S3) J. R. Lakovicz. *Principles of Fluorescence Spectroscopy*, 3rd ed.; Springer US: New York, 2006.
- (S4) Bi, S.; Ding, L.; Tian, Y.; Song, D.; Zhou, X.; Liu, X.; Zhang, H. Investigation of the Interaction between Flavonoids and Human Serum Albumin. *J. Mol. Struct.***2004**, 703 (1), 37–45.